GRUSONIA PULCHELLA CLASSIFICATION AND ITS IMPACTS ON THE GENUS GRUSONIA: MORPHOLOGICAL AND MOLECULAR EVIDENCE

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Summary. This report is an investigation into the relationships among members of the genus *Grusonia* sensu Anderson (2001) with an emphasis on examining the relationship of *Grusonia pulchella* within Opuntioideae (Cactaceae). *G. pulchella* is morphologically and geographically distinct from other *Grusonia* species, and nrITS DNA sequence data suggest that it may represent an independent evolutionary lineage from other grusonioid cacti. With the morphological, geographical, and molecular evidence considered, I propose to resurrect the generic concepts *Corynopuntia* Knuth (1935), *Grusonia* Reichenbach ex Schumann (sensu Britton and Rose, 1919), and *Micropuntia* Daston (1946).

Introduction

The current circumscription of the Grusonia pulchella (sand cholla) complex includes several described taxa. The first was Opuntia pulchella (Engelmann, 1863), whose type locality is on the Walker River in Nevada. Several specimens from Utah and bordering Nevada were described as new species under the new genus Micropuntia (Daston, 1946), i.e., Micropuntia barkleyana, M. brachyrhopalica, and M. spectatissima. Benson's (1957) circumscription of O. pulchella included these three morphotypes (as "aberrant forms") and notes that the species' affinities are difficult to ascertain. Robinson (1973) combined sand cholla along with seven other species into the genus Grusonia, previously described by Reichenbach (1896). The most recent treatments of Grusonia also include the genera Corynopuntia Knuth, Marenopuntia Backeberg, and Micropuntia Daston (Anderson, 2001; Stuppy, 2002; Wallace and Dickie, 2002; Gibson, ined.)

The *G. pulchella* (Engelm.) H. Rob. complex is a morphologically variable taxon consisting of scattered populations of small, teretestemmed opuntioid cacti found in the Great Basin of western North America (Benson, 1982; Kartesz, 1987) (Fig. 1). Although plants of the *G. pulchella* complex have long been considered to possess affinities with other grusonioid cacti (Engelmann, 1863; Britton and Rose, 1919; Benson, 1957, 1982; Anderson, 2001; Stuppy, 2002), aspects of the morphology, distribution, and habit of *G. pulchella* suggest that it may represent an evolutionary lineage distinct from other opuntioids, as first suggested by Daston (1946).



Figure 1. Habitat of *Grusonia pulchella*: view south across the Columbus salt marsh from the ghost town/alluvial fan of Columbus, Esmeralda County, Nevada, USA.



Figure 2. Grusonia pulchella in an alluvial fan above the Columbus salt marsh.

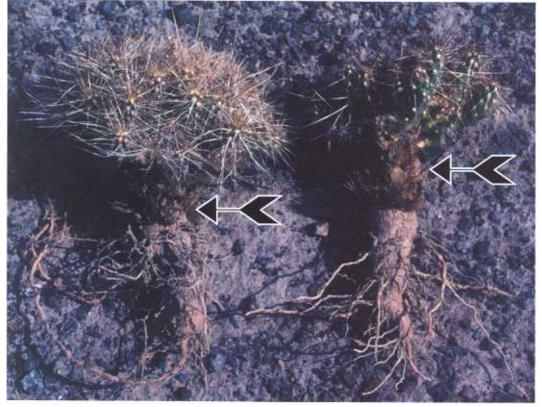


Figure 3. Two specimens of *Grusonia pulchella*. Note enlarged fleshy underground areole- and glochid-bearing structure.

Grusonia pulchella (Fig. 2) differs morphologically from other members of the genus in a number of significant ways. These plants are marked by an enlarged fleshy underground structure featuring areoles with prominent yellow glochids (Fig. 3). Some authors term this morphological feature a "tuberous root" (Benson, 1982; Anderson, 2001), but the areoles and glochids present upon the upper portion of the structure provide evidence that it may be stemderived (Britton and Rose, 1919) or perhaps a true tuber (Jackson, 1928). The presence of an underground tuber would make this species unique among the grusonioids. Other Grusonia species possess somewhat thickened roots (Engelmann, 1859; Benson, 1982; Ralston and Hilsenbeck, 1989), but these do not approach the proportions of the storage tuber of G. pulchella. They also are derived entirely from root tissue. Terminal stems of G. pulchella are often cylindrical rather than fully clavate (Britton and Rose, 1919) and are rather flexible, much like those of Pereskiopsis Britton & Rose, although much reduced in size. Unlike most Grusonia species, G. pulchella has tubercles that are often inconspicuous (Benson, 1982), but this varies (Britton and Rose, 1919; Daston, 1946; Anderson, 2001). The areoles of G. pulchella bear persistent wool similar to the areolar wool of *Pereskiopsis*, whereas the areoles of other Grusonia species have early-decidu-

ous wool or lack wool entirely (Anderson, 2001). Although displaying these subtle morphological affinities with Pereskiopsis, sand cholla obviously lacks the overall habit and persistent leaves of that genus. Spines on the fruit of G. pulchella differ from those of other Grusonia species by being antrorsely barbed rather than retrorsely barbed (Parfitt, 1988; Pinkava, 1999). Seeds of G. pulchella are sufficiently distinct morphologically from other Grusonia seeds as to place them in the monotypic subgenus Micropuntia sensu Stuppy (2002). One unique opinion even considers possible affinities of G. pulchella with Maihuenia (Philippi ex Weber) Schumann and Pterocactus Schumann (Daston, 1946).

Grusonia pulchella is disjunct from other members of the genus Grusonia (Fig. 4), which occur further to the southeast, through southernmost Nevada, Arizona, New Mexico, and Texas, and south into Mexico (Benson, 1982; Pinkava, 2002; Gibson, ined.). The geographically nearest congener of sand cholla is G. parishii (Orcutt)

Pinkava. Both species have been collected in Nye County, Nevada. Based on locality data from specimens cited in Benson (1982), the ranges of these two taxa do not appear to overlap. Furthermore, *G. partishit* occurs at elevations below 900-1200 m, and *G. pulchella* is found at 1200-1500 m and higher (Benson, 1969, 1982; Pinkava, 1999; Morefield, 2001). All chromosome counts of *G. pulchella* show it to be diploid, 2n = 22 (Pinkava, 2002).

The habit of *G. pulchella* differs from that of congeneric plants. The majority of *Grusonia* species are easily identifiable to genus at a distance, as they form dense, spreading mats of branching, clavate cladodes (Benson, 1982; Ralston and Hilsenbeck, 1989, 1992; Powell, 1998; de la Cerda-Lemus, 1999; Anderson, 2001), which sometimes root adventitiously (Engelmann, 1859). *Grusonia pulchella* is not matforming but is better described as a cushion or a dense, low-growing single-trunked shrub (Benson, 1982; Pinkava, 1999). The distal cladodes of *G. pulchella* do not root adventitiously while attached, nor do they detach easily (pers. obs.).

Recent molecular analyses have provided much insight into the relationships among the genera of Opuntioideae (Wallace and Dickie, 2002; Wallace and Gibson, 2002). Chloroplast DNA sequences demonstrate a sister group relationship of four species of *Grusonia* sensu

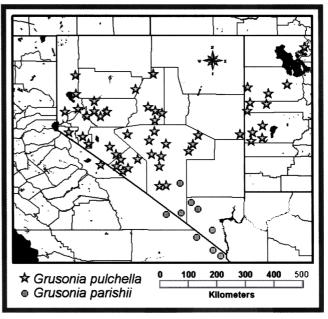


Figure 4. Map showing the distribution of *Grusonia pulchella* and its most geographically proximate congener, *G. parishii*. The range of *G. parishii* extends further south into California. Other *Grusonia* species occur further south and east, through the southwestern United States and into Mexico. Adapted from Benson, 1982; Albee et al., 1988; Morefield, 2001; Pinkava, 2002; and Gibson, ined.

Anderson (2001) with a limited sampling of Cylindropuntia (Engelm.) Knuth. Sampled Grusonia in these studies include the type species G. bradtiana, along with G. clavata, G. marenae, and G. stanleyi. These two genera form a well-supported monophyletic group with Pereskiopsis and Quiabentia Britton & Rose. The relationship of G. pulchella within the Opuntioideae has not yet been elucidated. Given the conflicting evidence of morphological data towards the relationship of G. pulchella within the Opuntioideae, there is a potential for molecular data to help resolve this species' taxonomic position. Toward this goal, I have gathered DNA sequence data from the nrITS region for specimens of G. pulchella and putative related taxa.

Methods and Materials

Live material was obtained through fieldwork or from live vouchered plantings at Ran-

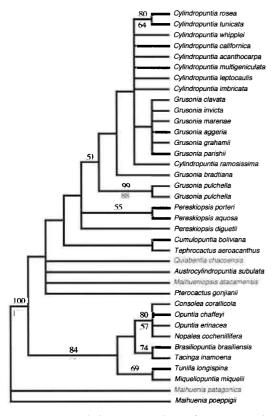


Figure 5. A phylogenetic analysis of *Grusonia pulchella* and related taxa. Strict consensus of 809,300 most-parsimonious trees found by the fleuristic search of the nrITS DNA data. With uninformative characters excluded, length = 208; CI = 0.6667; RI = 0.8427; rescaled consistency = 0.6847. Bootstrap percentages above 50% are indicated above the branches, and jackknife support above 50% is indicated below the branches.

cho Santa Ana Botanic Garden, Claremont, CA; Desert Botanical Garden, Phoenix, AZ; and Sul Ross State University Cactus Garden, Alpine, TX. This study sampled 34 specimens, including 2 specimens of G. pulchella, 7 specimens of other *Grusonia* species, including representatives from all other described genera currently circumscribed within Grusonia sensu Anderson (2001), and 25 specimens representing all other genera currently recognized in Opuntioideae (Table 1). DNA was extracted using 2X CTAB, followed by precipitation in cold isopropanol (Griffith and Porter, in press). All extractions used approximately 0.5 g of epidermal shoot tissue. Polymerase Chain Reaction (PCR) amplification of templates of the nuclear ribosomal internal transcribed spacer region (nrITS) follows the methods outlined by Columbus et al. (1998) using the primers ITS4 and ITS5 (White et al., 1990). Polyethylene glycol precipitation (Morgan and Soltis, 1993) purified all templates. Purified template amplifications were sequenced directly with four primers, ITS2, ITS3, ITS4i, and ITS5i (White et al., 1990; Porter, 1997), using "big dye" chemistry from Applied Biosystems Incorporated, according to the manufacturer's specifications. An Applied Biosystems Incorporated 3100 automated DNA sequencer gathered all sequences from PCR products. Genbank accessions from a previous study (Hershkovitz and Zimmer, 1997) provided 3 additional sequences representing outgroup taxa.

Chromatograms from sequencing reactions were assembled into contigs and edited using Sequencher v4.1 (Gene Codes Corporation, Inc.). Consensus sequences were aligned manually with Se-Al v2.0a72 (Rambaut, 1996). Gaps were treated as missing data. Informative indels were coded. Phylogenetic relationships among these taxa were estimated using Fitch parsimony, in PAUP* v4.0ß8 (Swofford, 1998). Estimations of confidence in the clades obtained by these analyses were gathered through bootstrap analysis (Felsenstein, 1985) with 10,000 pseudoreplicates, and through jackknifing (Farris et al., 1996), also with 10,000 pseudoreplicates (63% deletion) as performed by PAUP*.

Results and Discussion

As a whole, *Grusonia* sensu Anderson (2001) is not supported as a monophyletic group, rather, it is apparently paraphyletic. The two specimens of *G. pulchella* form a well-supported monophyletic group (99% bootstrap, 88% jackknife) that is sister to the remaining samples of *Grusonia* (*Corynopuntia*, *Grusonia*, and *Marenopuntia*) and all sampled members of the genus *Cylindropuntia* (Fig. 5). Although this sister group relationship lacks strong statistical support, this topology was recovered in all 809,300 most-parsimonious

Table 1. Sources of nrITS DNA sequences for specimens used in molecular analysis

Species	Source*	Voucher
Austrocylindropuntia subulata (Muehlenpfordt) Backeberg	DBG 1990 0692	
Brasiliopuntia brasiliensis (Schumann) Berger	DBG 1990 0559	
Consolea corallicola Small	DBG 1997 0397	
Cumulopuntia boliviana (Salm-Dyck) F. Ritter	DBG 1970 9884	
Cylindropuntia acanthocarpa (Engelm. & Bigelow)	San Bernardino County, CA, USA	Griffith 211 (RSA)
Cylindropuntia echinocarpa (Engelm. & Bigelow) Knuth	Clark County, NV, USA	Griffith 200 (RSA)
Cylindropuntia imbricata (Haworth) Knuth	Nuevo León, Mexico	Griffith 250 (RSA)
Cylindropuntia leptocaulis (de Candolle) Knuth	Coahuila, Mexico	Griffith 244 (RSA)
Cylindropuntia californica (J. Torrey & Gray) Knuth	Los Angeles County, CA, USA	Columbus s.n. (RSA
Cylindropuntia ramosissima (Engelm.) Knuth	Clark County, NV, USA	Griffith 202_(RSA)
Cylindropuntia rosea (de Candolle) Backeberg	Puebla, Mexico	Griffith 187_(RSA)
Cylindropuntia tunicata (Lehmann) Knuth	Coahuila, Mexico	Griffith 256 (RSA)
Cylindropuntia wbipplei (Engelm. & Bigelow) Knuth	San Juan County, NM, USA	Porter s.n. (RSA)
Grusonia aggeria (Ralston and Hilsenbeck) Anderson	SRSU	Powell 6006 (SRSC)
Grusonia bradtiana (Coulter) Britton & Rose	DBG 1985 0345	
Grusonia invicta (Brandegee) Anderson	RSA	Griffith 218 (RSA)
Grusonia grahamii (Engelm.) H. Rob.	SRSU	Hardy 634 (SRSC)
Grusonia marenae (Parsons) Anderson	DBG 1954 4980	
Grusonia parishii (Orcutt) Pinkava	RSA	Wisura s.n. (RSA)
Grusonia pulchella(Engelm.) H. Rob.	Churchill County, NV, USA	Griffith 210 (RSA)
	Esmeralda County, NV, USA	Griffith 353 (RSA)
Grusonia villis (Rose) H. Rob.	Coahuila, Mexico	Griffith 246 (RSA)
Maihuenia patagonica (Philippi) Britt. & Rose	Hershkovitz and Zimmer, 1997	
Maihuenia poeppigii (Otto ex Pfeiffer) Philippi ex Schumann	Hershkovitz and Zimmer, 1997	
Maihueniopsis atacamensis (Philippi) Ritter	DBG 2001 0112	
Miqueliopuntia miquelii (Monville) F. Ritter	DBG 1997 0129	
Nopalea cochenillifera (L.) Salm-Dyck	DBG 1997 0395	
Opuntia chaffeyi Britt. & Rose	D B G 1990 0238	
Opuntia erinacea Engelm. & Bigelow	Mono County, CA, USA	Honer 658 (RSA)
Pereskiopsis aquosa (Weber) Britt. & Rose	DBG 1997 0001	
Pereskiopsis porteri (Brande. ex Weber) Britt. & Rose	Hershkovitz and Zimmer, 1997	
Pereskiopsis diguettii (Weber) Britt. & Rose	Michoacan, Mexico	Griffith 169 (RSA)
Pterocactus gonjianii Kiesling	DBGs.n.	
Quiabentia chacoensis Backeberg	DBG 1985 0046	
Tacinga inamoena (Schumann) Stuppy & Taylor	DBG 1999 0017	
Tephrocactus aeroacanthus (Lemaire) Lemaire	DBG 2001 0115	
Tunilla corrugata (Salm-Dyck) Hunt & Iliff	D B G Hunt 66371	

^{*}D B G = Desert Botanical Garden, 1201 North Galvan Parkway, Phoenix, Arizona, USA; R S A = Rancho Santa Ana Botanic Garden, 1500 North College Avenue, Claremont, California, USA; S R S U = Sul Ross State University Cactus Garden, Alpine, Texas, USA.

trees. In addition, the type species of Grusonia (G. bradtiana) is sister to all sampled species of Cylindropuntia and Grusonia, with the exception of G. pulchella. The remaining Grusonia species sampled form a monophyletic group, although this clade also lacks strong statistical support. Since the paraphyly of Grusonia sensu Anderson (2001) lacks such support, hypotheses that propose an affinity between G. pulchella and other grusonioid cacti (Engelmann, 1863; Britton and Rose, 1919; Benson, 1957, 1982; Anderson, 2001; Stuppy, 2002) cannot be rejected by these data. Although the relationships between Cylindropuntia and Grusonia also lack strong statistical support, these data provide evidence that Cylindropuntia is nested within Grusonia sensu Anderson (2001). The current circumscription of Grusonia (Anderson, 2001; Stuppy, 2002) therefore taxonomically recognizes a non-monophyletic group.

Although the ITS data do resolve differences between Cylindropuntia and Grusonia, the sister group relationship of G. pulchella to all other non-tropical North American teretestemmed opuntioids may indicate a distinct evolutionary lineage for the sand cholla. In the context of recent tendencies to split the large genus Opuntia sensu Benson (1982) into smaller genera reflecting natural groups (Robinson, 1973; Anderson, 1999, 2001; Stuppy, 2002; Wallace and Dickie, 2002), I propose a solution to improve the taxonomy of the grusonioid cacti. In the context of the morphological evidence, geographical separation, and the monophyly of the G. pulchella lineage, the recognition of these plants under the genus Micropuntia is supported. The generic concept of Grusonia may revert to that of Britton and Rose (1919), circumscribing only the type species G. bradtiana. Support for this narrow concept can also be found in that species' morphological distinctness from other Grusonia species (Britton and Rose, 1919; Anderson, 2001; Stuppy, 2002) and in the monophyly of the remaining species. These remaining species of Grusonia sensu Anderson (2001) should be treated under the genus Corynopuntia sensu Knuth (1935). including G. marenae (Parsons) Anderson (but excluding G. pulchella). Morphological support for this generic concept exists in the form of seed characters used to circumscribe Grusonia subg. Corynopuntia sensu Stuppy (2002), in addition to the characters discussed above. A summary of the generic circumscriptions proposed here is presented in Table 2. The other solution would be to include Corynopuntia, Cylindropuntia, Marenopuntia, and Micropuntia into a broadly circumscribed Grusonia (which has priority). I am not in favor of this option, although this broad circumscription would certainly recognize a monophyletic

group. However, it would unnecessarily obscure the natural morphological diversity of the lineage as well as the morphological cohesiveness of many of its members.

I am aware of the lack of strong statistical support for many of the relationships suggested by the DNA sequence data. However, as there is even less support for the monophyly of *Grusonia* sensu Anderson (2001), I propose the recognition of these older generic concepts. Ongoing investigations involving additional DNA sequencing of nuclear and chloroplast genes and anatomical work may further elucidate these relationships, allowing for the most perceptive taxonomy possible. Further investigations may also elucidate whether specific rank is warranted for the various morphotypes (Daston, 1946; Benson, 1957) currently circumscribed within *Micropuntia pulchella*.

Nomenclatural Changes

The following new combinations are needed in support of this manuscript:

Corynopuntia aggeria (Ralston and Hilsenbeck) M. P. Griffith, comb. nov. Basionym: Opuntia aggeria Ralston and Hilsenbeck, Madroño 4: 226. 1989.

Corynopuntia emoryi (Engelm.) M.P. Griffith, comb. nov. Basionym: Opuntia emoryi Engelm., Proc. Am. Acad. 3: 303. 1857.

Corynopuntia kunzei (Rose) M.P. Griffith, comb. nov. Basionym: Opuntia kunzei Rose, Smithsonian Misc. Collect. 50: 505. 1908.

Corynopuntia marenae (Parsons) M.P. Griffith, comb. nov. Basionym: Opuntia marenae Parsons, Desert Pl. Life 8: 10. 1936

Micropuntia pulchella (Engelm.) M.P. Griffith, comb. nov. Basionym: Opuntia pulchella Engelm., Trans. Acad. Sci. St. Louis 2: 201. 1863.

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Table 2. Summary of generic circumscriptions presented here

Genus	Included species	Synapomorphies, Geography	
Corynopuntia Knuth	C. aggeria (Ralston & Hilsenbeck) M.P. Griffith	ith Mat-forming habit with growth to 50 cm (60 cm in <i>C. marenae</i>) composed of clavate segments (terete in <i>C. marenae</i>). Spines of fruits retrorsely barbed, stiff. Deserts and grasslands of Northern Mexico and Southwestern United States to Nye County, Nevada.	
c. c. c. c. c. c. c. c.	C. agglomerata (Berger) Knuth		
	C. bulbispina (Engelm.) Knuth		
	C. clavata (Engelm.) Knuth		
	C. dumetorum (Berger) Knuth		
	C. emoryi (Engelm.) M.P. Griffith		
	C. grahamii (Engelm.) Knuth		
	C. invicta (Brandegee) Knuth		
	C. kunzei (Rose) M.P. Griffith		
	C. marenae (Parsons) M.P. Griffith		
	C. moellerina Knuth		
	C. parishii (Orcutt) Knuth		
	C. reflexispina (Wiggins & Rollins) Backeberg		
	C. schottii (Engelm.) Knuth		
	C. vilis (Rose) H. Robinson		
Grusonia Reichenbach	G. bradtiana (Coulter) Britton & Rose	Mat-forming habit with erect, ribbed, jointed growth to 1 m. Flowering areoles without glochids. Chihuahuan Desert of Coahuila, Mexico.	
Micropuntia Daston	M. pulchella (Engelm.) M.P. Griffith	Cushion habit to 20 cm high; storage tuber with areoles, glochids. Spines of fruit antrorsely barbed, flexible. Great Basin of Nevada, Utah, and California, United States.	

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Literature Cited

- Albee, B.J., L.M. Schultz, and S. Goodrich. 1988. Atlas of the vascular plants of Utah. Utah Museum of Natural History, Salt Lake City.
- Anderson, E.F. 1999. Some nomenclatural changes in the Cactaceae subfamily Opuntioideae. Cact. Succ. J. (US) 71: 324-325
- ———. 2001. The cactus family. Timber Press, Portland, Oregon.
- Benson, L.H. 1957. The *Opuntia pulchella* complex. Cact. Succ. J. 29: 19-21.
- 1969. The native cacti of California. Stanford University Press, Stanford.
- ——. 1982. The cacti of the United States and Canada. Stanford University Press, Stanford.
- Britton, N.L., and J.N. Rose. 1919. The Cactaceae. Volume 1. Carnegie Institution, Washington, D.C.
- Columbus, J.T., M.S. Kinney, R. Pant, and M.E.

- Siqueiros Delgado. 1998. Cladistic parsimony analysis of internal transcribed spacer region (nrDNA) sequences of *Bouteloua* and relatives (Gramineae: Chloridoideae). Aliso 17: 99-130.
- Daston, J.S. 1946. Three noteworthy cacti of southwestern Utah. Amer. Midl. Nat. 36: 661-662.
- De la Cerda-Lemus, M. 1999. Cactáceas de Aguascalientes. Universidad Autónoma de Aguascalientes, Mexico.
- Engelmann, G. 1859. Cactaceae of the Boundary. In
 W.H. Emory. 1859. Report of the United States and
 Mexican Boundary Survey. Volume 2. A.O.P.
 Nicholson, Washington.
- Farris, J.S., V.A. Albert, M. Kallersjo, D. Lipscomb, and A.G. Kluge. 1996. Parsimony jackknifing outperforms neighbor-joining. Cladistics 12: 99-124.
- Felsenstein, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. Evolution 39:783-791.
- Gibson, A., ined. Cactaceae. *In* Flora of North America Editorial Committee, ed. Flora of North America. Oxford University Press, New York.
- Griffith, M. P., and J. M. Porter. In press. Back to the ba-

- sics: a simple method of DNA extraction for mucilaginous cacti. Bradleya 21.
- Hershkovitz, M.A., and E.A. Zimmer. 1997. On the evolutionary origins of the cacti. Taxon 46: 217-232.
- Jackson, B.D. 1928. A glossary of botanic terms. 4th edition. Phototype Limited, London.
- Kartesz, J.T. 1987. A flora of Nevada. Unpublished Ph.D. dissertation, University of Nevada, Reno.
- Knuth, Count F.M. 1935. Corynopuntia. In C. Backeberg and Count F.M. Knuth. 1935. Kaktus-ABC. Gyldendals Forlagstrykkeri, Copenhagen.
- Morefield, J.D. (editor). 2001. Nevada Rare Plant Atlas. Carson City: Nevada Natural Heritage Program, compiled for the U.S. Department of Interior, Fish and Wildlife Service, Portland, Oregon and Reno, Nevada.
- Morgan, D.R., and D.E. Soltis. 1993. Phylogenetic relationships among members of Saxifragaceae sensu lato based on *rbcL* sequence data. Ann. Miss. Bot. Gard. 80: 631-660.
- Parfitt, B.D. 1988. *Opuntia clavata* and *Opuntia pul-chella*: present or absent in Arizona? Cact. Succ. J. (US) 60: 227-229.
- Pinkava, D.J. 1999. Cactaceae: Cactus Family, Part Four, Grusonia. J. Arizona-Nevada. Acad. Sci. 32: 48–52.
- ——. 2002. On the evolution of continental North American Opuntioideae Succ. Pl. Res. 6: 59-98.
- Porter, J.M. 1997. Phylogeny of Polemoniaceae based on nuclear ribosomal internal transcribed spacer DNA sequences. Aliso 15: 57-77.
- Powell, A.M. 1998. Trees and shrubs of Trans-Pecos Texas and adjacent areas. University of Texas Press, Austin.

- Ralston, B.E., and R.A. Hilsenbeck. 1989. Taxonomy of the *Opuntia schottii* complex (Cactaceae) in Texas. Madroño 36: 221-231.
- ——. 1992. Opuntia densispina (Cactaceae): a new club cholla from the Big Bend region of Texas. Madroño 39: 281-284.
- Rambaut, A. 1996. Se-Al. Sequence alignment editor. Version 2.0a7.2. Department of Zoology, University of Oxford.
- Reichenbach, F. 1896. *Grusonia. In* Schumann, K. 1896. Monats. Kakteenk. 6: 177.
- Robinson, H. 1973. New combinations in the Cactaceae subfamily Opuntioideae. Phytologia 26: 175-176.
- Stuppy, W. 2002. Seed characters and generic classification of Opuntioideae. Succ. Pl. Res. 6: 25-58.
- Swofford, D.L. 1998. PAUP*. Phylogenetic Analysis Using Parsimony (*and Other Methods). Version 4. Sinauer Associates, Sunderland, Mass.
- Wallace, R.S., and S.L. Dickie. 2002. Systematic implication of chloroplast DNA sequence variation in subfam. Opuntioideae (Cactaceae). Succ. Pl. Res. 6:9-24.
- Wallace, R.S., and A.C. Gibson. 2002. Evolution and systematics. *In P.S. Nobel* (ed.). Cacti: biology and uses. University of California Press, Berkeley.
- White, T. J., T. Bruns, S. Lee, and J. Taylor. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *In M.A. Innis, D.H. Gelfand, J.J. Sninsky, and T.J. White (eds.)*. PCR protocols: a guide to methods and applications. Academic Press, San Diego.